Antibiotic Susceptibility Prediction Using JNN

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Abstract: In this research, an Artificial Neural Network (ANN) model was developed and validated to predict efficiency of antibiotics in treating various bacteria types. Attributes that were taken in account are: organism name, specimen type, and antibiotic name as input and susceptibility as an output. A model based on one input layer, one hidden layer, and one output layer concept topology was developed and trained using a data from Queensland government's website. The evaluation shows that the proposed ANN model using JNN tool is capable of correctly predicting the susceptibility of organisms to the antibiotics with 94.17% accuracy.

Keywords: Artificial Neural Networks, antibiotic susceptibility, ANN, JNN

1. INTRODUCTION

An antibiotic is a chemical produced by living organisms to protect themselves from attacking bacteria or viruses. People made use of antibiotics in fighting infectious diseases. Many antibiotics nowadays are synthetic or semisynthetic. Bacteria respond to antibiotics in different ways, i.e. they are resistive or sensible or moderate sensible to a particular antibiotic. There are many methods to test the antibiotic susceptibility. The most famous one is the Kirby-Bauer test (or the disc diffusion test). But these methods may last from few hours to many days until they yield the results. To make things easier we seek to computerize antibiotic susceptibility test.

This study seeks to automate the measure process of antibiotic susceptibility using artificial neural network technology.

We used the JustNN tool to train the ANN on the "antibiogram-2013" dataset provided by Queensland government website: www.qld.gov.au.

The objective of this study is: to make the antibiotic susceptibility test easy for physicians and scientists,

2. LITERATURE REVIEW

a) Michael Hombach and others made a study to assess the rapid fully automated disc diffusion for rapid antibiotic susceptibility test results. They claimed: "We have evaluated a fully automated system for its potential for early reading of disc delusion diameters after 6–12h of incubation. We assessed availability of results, methodological precision, categorical agreement, and interpretation errors as compared with an 18h standard. In total, 1028 clinical strains (291 Escherichia coli, 272 Klebsiella pneumoniae, 176 Staphylococcus aureus and 289 Staphylococcus epidermidis) were included in this study. Disc delusion plates were streaked, incubated and imaged using the WASPLab automation system"[1].

- b) Kiran Desai proved that "support vector regression (SVR)" is an attractive alternative to artificial neural networks for the development of soft-sensors in bioprocesses [2].
- c) I.A Basheer et al. praised ANN technology saying: "The attractiveness of ANNs comes from their remarkable information processing characteristics pertinent mainly to nonlinearity, high parallelism, fault and noise tolerance, and learning and generalization capabilities. This paper aims to familiarize the reader with ANN-based computing (neurocomputing) and to serve as a useful companion practical guide and toolkit for the ANNs modeler along the course of ANN project development. The history of the evolution of neurocomputing and its relation to the field of neurobiology is briefly discussed. ANNs are compared to both expert systems and statistical regression and their advantages and limitations are outlined. A bird's eye review of the various types of ANNs and the related learning rules is presented, with special emphasis on backpropagation (BP) ANNs theory and design. A generalized methodology for developing successful ANNs projects from conceptualization, to design, to implementation, is described. The most common problems that BPANNs developers face during training are summarized in conjunction with possible causes and remedies. Finally, as a practical application, BPANNs were used to model the microbial growth curves of S. flexneri. The developed model was reasonably accurate in simulating both training and test time-dependent growth curves as affected by temperature and pH" [3].
- d) Guoqiang Zhang et al. present a state-of-the-art survey of ANN applications in forecasting. The purpose of the paper is to provide (1) a synthesis of published research in this area, (2) insights on ANN modeling issues, and (3) the future research directions [6].
- e) Sovan Lek et al. made a good paper to present some of the most important papers of the first workshop about ANNs in ecological modeling [4].

f) M.W Gardner et al. wrote a paper to present a general introduction and discussion of recent applications of the multilayer perceptron, one type of artificial neural network, in the atmospheric sciences [5].

3. ARTIFICIAL NEURAL NETWORKS

Artificial Neural Networks (ANN) are tiling the way for life-shifting applications to be developed for use in all areas of the economy. Artificial Intelligence (AI) podiums that are built on ANN are distracting the traditional way of doing things [6]. From interpreting web pages into other languages to taking virtual assistant order groceries online to chatting with chatbots to resolve problems, AI podiums are simplifying transactions and making facilities reachable to all at small costs.

ANN are built similar to the human brain [7], with neuron nodes connected like a web. The human brain has billions of cells called neurons. Each neuron is consist of a cell body that is accountable for processing information by carrying information as inputs giving out outputs from the brain. ANN has thousands of artificial neurons called processing units, which are connected by nodes [8]. These processing units are consists of input and output units. The input units receive numerous forms and structures of information based on an interior weighting system and the neural network tries to learn about the information accessible in order to create one output report[9]. As humans need rules and guidelines to produce a result, ANNs likewise use a set of learning rules called backpropagation, an abbreviation for backwards propagation of error, to faultless their output results [10].

An ANN firstly goes through a training stage where it learns to identify patterns in data, whether visually, aurally or textually. Throughout this supervised stage, the network matches its actual output produced with what it was meant to produce, i.e. the wanted output. The difference among both results is attuned using backpropagation. This means that the network works backwards going from the output unit to the input units to be able to amend the weight of its connections among the units till the difference among the real and wanted result yields the lowest potential error[11].

ANN have been practical in all areas of operations. Email service suppliers use ANN to discover and delete spam from a user's inbox; help managers to forecast the direction of a company's stock; Credit rating firms use it to enhance their credit scoring approaches; e-commerce platforms use it to personalize recommendations to their customers; chatbots are developed with ANN for natural language processing; deep learning algorithms use ANN to predict the likelihood of an incident; and the list of ANN incorporation goes on across multiple areas, businesses and countries[12].

4. METHODOLOGY

4.1 Search for appropriate dataset.

Antimicrobial susceptibility data extracted from the Queensland Health Pathology reporting system. The data provide information about the level of resistance certain organisms have to antimicrobial agents typically used to treat infections. The "antibiogram-2013" dataset contains 358 records with 5 attributes: "organism", "specimen type", "antibiotic", "% susceptible", and "number of isolates tested". It assigns a percentage of antibiotic-susceptibility to each organism in blood, urine, and other specimens, (e.g. the organism "Streptococcus pneumonia" that is found in "blood" specimen is 91% susceptible to the

4.2 Normalize output column in the dataset.

In this step we didn't alter the attributes arrangement but we applied the normalization formula:

"erythromycin" antibiotic. This result was calculated after 178 isolates were tested)[14].

$$Xi = \frac{X \max - Xi}{X \max - X \min}$$

on the output attribute "% susceptible".

No.	Attributes	Data type	Chosen as	
1.	organism	Text	Input	
2.	specimen type	Text	Input	
3.	antibiotic	Text	Input	

Table 1:	Attributes	of the	Dataset
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4.	% susceptible	Real	Output
5.	isolates tested	Integer	Input

4.4 Building the ANN Model

We have used Just Neural Network (JNN) tool [15] to build a multilayer ANN model. The first step after normalizing the data, it was imported into JNN environment (As in Figure 1). We have sat the parameters of the proposed model as follows: Learning Rate 0.39 and the Momentum to be 0.51, and Average Error rate to be 0.01 (as shown in Figure 2).

The proposed model consists of 3 Layers: Input Layer with 4 nodes, the Hidden Layer with 5 nodes, and Output Layer with one node as can be seen in Figure 3.

4.5 Evaluating the ANN model

The Antibiogram dataset consists of 358 samples with 5 attributes as in Table 1. We divided the imported dataset into two groups (Training and Validation) randomly using the JNN tool. The Training consists of approximately 67% (238 samples) and the validation set consists of 33% of the dataset (120 samples). After making sure that the parameter control was set properly, we started training the ANN model and kept eye on the learning curve, loss error and validation accuracy. We trained the ANN model for 2481 cycles. The best accuracy we got was 94.17% (as seen in Figure 4). We determined the most influential factors in the Antibiogram dataset as in Figure 5. Figure 6 shows the summary of the proposed model.

The following figure shows the architecture of the neural network with one input layer, one hidden layer (contains four neurons), and one output layer.

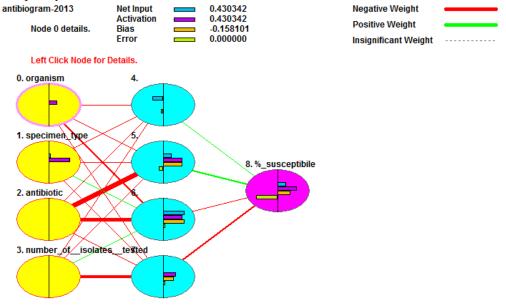


Figure 3: Final ANN model design.

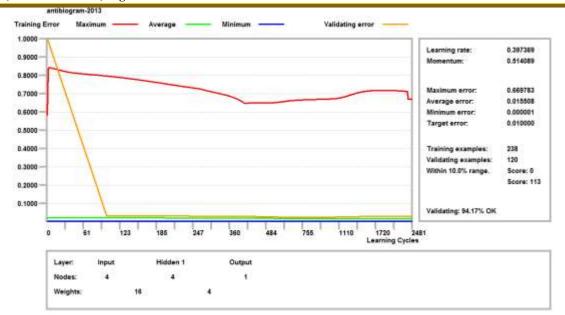


Figure 4: ANN model raining and validation

Controls	
Learning Learning rate 0.39736: ✓ Decay ✓ Optimize Momentum 0.51408: ✓ Decay ✓ Optimize	Target error stops
Validating Cycles <u>b</u> efore first validating cycle 100 Cycles per validating cycle 100	Validating stops Image: Ima
Select O examples at random from the Training examples = 238	or Correct after rounding Fixed period stops
Slow learning Delay learning cycles by 0 millisecs	Stop after 20.0000 seconds Stop on 0 cycles
	OK Cancel

Figure 2: Setting Parameters of the proposed model

antibiogram-2013 2481 cycles. Target error 0.0100 Average training error 0.015508 The first 4 of 4 Inputs in descending order.

Column	Input Name	Importance	Relative Importance
2 4 0 1	antibiotic number_ofisolates organism specimen_type	36.8338 	

Figure 5: Most influential filed in the dataset

	organism	specimen_t+	antibiotic	<pre>%_suscepti+</pre>	number_of_+	
ŧ0	Escherichia	0	amikacin	0.9900	0.0628	
ŧ1	Klebsiella	0	amikacin	1.0000	0.0143	
2	Pseudomonas	0	amikacin	0.9900	0.0102	
±3	Enterobacte	0	amikacin	0.9900	0.0057	
ŧ4	Proteus mir	0	amikacin	0.9900	0.0027	
ŧ5	Serratia ma	0	amikacin	1.0000	0.0024	
ŧ6	Klebsiella	0	amikacin	1.0000	0.0015	
ŧ7	Enterobacte	0	amikacin	1.0000	0.0006	
ŧ8	Acinetobact	0	amikacin	1.0000	0.0006	
ŧ9	Citrobacter	0	amikacin	1.0000	0.0005	
ŧ10	Pseudomonas	2	amikacin	0.9000	0.2149	
ŧ11	ESCAPPM gro	2	amikacin	0.9700	0.0869	
ŧ12	Escherichia	2	amikacin	0.9900	0.0646	
‡13	Klebsiella	2	amikacin	1.0000	0.0303	
ŧ14	Proteus sp.	2	amikacin	1.0000	0.0141	
ŧ15	Acinetobact	2	amikacin	0.9900	0.0130	
‡16	Klebsiella	2	amikacin	1.0000	0.0085	
ŧ17	Citrobacter	2	amikacin	1.0000	0.0038	
‡18	Escherichia	1	amikacin	1.0000	0.8967	
‡19	Klebsiella	1	amikacin	1.0000	0.1460	
ŧ20	Pseudomonas	1	amikacin	0.9600	0.1218	
21	Proteus mir	1	amikacin	1.0000	0.0838	
‡22	Enterobacte	1	amikacin	0.9900	0.0592	
23	Citrobacter	1	amikacin	1.0000	0.0207	
\$24	Klebsiella	1	amikacin	1.0000	0.0200	
ŧ25	Morganella	1	amikacin	0.9900	0.0170	
26	Serratia sp	1	amikacin	0.9900	0.0159	

Figure 1: Imported Normalized dataset to JNN environment

Details of antibiogram-2013		X			
General antibiogram-2013					
Learning cycles: 2481		AutoSave cycles: 100			
Training error: 0.015508		Validating error: 0.026755			
Validating results: 94.17% co	orrect after	rounding.			
Grid		Network			
Input columns: 4 Output columns: 1	i	Input nodes connected: 4			
Excluded columns: C)	Hidden layer 1 nodes: 4 Hidden layer 2 nodes: 0			
	238	Hidden layer 3 nodes: 0			
Validating example rows: 1 Querying example rows: C Excluded example rows: C Duplicated example rows: C)	Output nodes: 1			
Controls					
Learning rate: 0.	.3974	Momentum: 0.5141			
Validating 'correct' target: 1	00.00%				
Target error: 0.	.0100	Decay.			
Validating rules		Missing data action			
No columns have rules se	xt.	The median value is used.			
✓ Show when a file is opened					
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Figure 6: Detail of the proposed Model

www.ijeais.org/ijaisr

5. CONCLUSION

An artificial Neural Network model for predicating Antibiogram dataset was presented. The model used feed forward backpropagation algorithm for training the proposed ANN model using JNN tool. The factors for the model were obtained from dataset which represents Antibiogram dataset. The model was tested and the accuracy rate was 94.17%. This study showed that artificial neural network is capable of predicating Antibiogram dataset accurately.

References

- 1. Journal of Antimicrobial Chemotherapy, Volume 72, Issue 6, 1 June 2017, P ages 1659–1668.
- 2. Kiran, D. et al.(2006). "Soft-sensor development for fed-batch bioreactors using support vector regression." Biochemical Engineering Journal, 27(3).
- 3. Sovan, L. (1999). "Artificial neural networks as a tool in ecological modeling, an introduction." Ecological Modeling, 120(3).
- 4. Gardner, M.W. & Dorling, S.R. (1998). "Artificial neural networks (the multilayer perceptron)—a review of applications in the atmospheric sciences." Atmospheric Environment, 32(15), 2627-2636.
- 5. Basheer, M Hajmeer, Artificial neural networks: fundamentals, computing, design, and application, Journal of Microbiological Methods, Volume 43, Issue 1, 2000, Pages 3-31.
- 6. Guoqiang Zhang, B. Eddy Patuwo, Michael Y. Hu, Forecasting with artificial neural networks:: The state of the art, International Journal of Forecasting, Volume 14, Issue 1, 1998, Pages 35-62.
- 7. Atzberger, C. 2004. Object- based retrieval of biophysical canopy variables using artificial neural nets and radiative transfer models.. Remote Sensing of Environment, 93: 53–67.
- Augusteijn, M. F., Clemens, L. E. and Shaw, K. A. 1995. Performance evaluation of texture measures for ground cover identification in satellite images by means of a neural network classifier.. IEEE Transactions on Geoscience and Remote Sensing, 33: 616–626.
- 9. Augusteijn, M. F. and Folkert, B. A. 2002. Neural network classification and novelty detection.. International Journal of Remote Sensing, 23: 2891–2902.
- 10. Augusteijn, M. F. and Warrender, C. E. 1998. Wetland classification using optical and radar data and neural network classification.. International Journal of Remote Sensing, 19: 1545–1560.
- 11. Azimi- Sadjadi, M. R., Ghaloum, S. and Zoughi, R. 1993. Terrain classification in SAR images using principal components analysis and neural networks.. IEEE Transactions on Geoscience and Remote Sensing, 31: 511–515
- 12. Bandyopadhyay, S. 2005. Satellite image classification using genetically guided fuzzy clustering with spatial information. International Journal of Remote Sensing, 26: 579–593.
- Baraldi, A., Binaghi, E., Blonda, P., Brivio, P. A. and Rampini, A. 2001. Comparison of the multilayer perceptron with neurofuzzy techniques in the estimation of cover class mixture in remotely sensed data.. IEEE Transactions on Geoscience and Remote Sensing, 39: 994–1005.
- 14. Queensland government website: www.qld.gov.au.
- 15. EasyNN Tool